Identification of Three New Pterocarpans (6a,11a-Dihydro-6H-benzofuro[3,2-c][1]benzopyrans) from *Pisum sativum* infected with *Fusarium solani* f. sp. *pisi*

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The isolation of three antifungal pterocarpans from Pisum sativum epicotyls infected with Fusarium solani f. sp. pisi is described. By a combination of spectroscopic and degradative techniques the compounds were identified as 4-hydroxy-2,3,9-trimethoxy- (1), 3-hydroxy-2,9-dimethoxy- (2), and 2,3,9-trimethoxy-pterocarpan (3).

THE theory that plants accumulate antifungal compounds called phytoalexins in response to challenge by fungal pathogens was first formulated by Müller and Börger.¹ Phytoalexins produced by plants of the family Leguminosae often have the pterocarpan ring structure as in (1).² Previous investigations have demonstrated that garden pea (Pisum sativum L.) 6a-hydroxy-3-methoxy-8,9-methylenedioxyproduces pterocarpan (pisatin)^{2b} and 3-hydroxy-8,9-methylenedioxypterocarpan (maackiain)^{2c} in response to fungal challenge. We now report the identification of three additional pterocarpans from pea epicotyls infected with the fungus Fusarium solani (Mart.) Sacc. f. sp. pisi (F. R. Jones) Snyd. & Hans.

The mass spectra of the compounds, in which the molecular ions are the base peaks but which show little further fragmentation, are like those reported for pterocarpans.^{3,4} The n.m.r. spectrum of each compound contains the complex four-spin system which arises from the heterocyclic protons of pterocarpans.⁵ Carbonyl stretching absorbance is absent in the i.r. spectrum, but several prominent bands characteristic of aryl ether functionality are present.⁶ All three compounds gave a negative Labat reaction,⁷ which indicates the absence of methylenedioxy-groups.

Compound (1), $C_{18}H_{18}O_6$ (*M*⁺ 330), is phenolic, as evidenced by positive reaction with FeCl₃ and diazotized p-nitroaniline. Hydroxy-stretching absorbance is present in the i.r. spectrum. Reaction with diazomethane afforded a monomethylated product $(M^+ 344)$, and reaction with acetic anhydride produced a monoacetylated product $(M^+ 372)$. Neither product retained phenolic character. The n.m.r. spectrum of compound (1) contains the signals of three methoxy-groups and one hydroxy-group.

Distinction between ring-A and ring-D substituents of pterocarpans cannot be made on the basis of mass spectra, because any fragment ion may reasonably arise from either ring.³ However, Nakayama et al.⁴ have

m/e 148 (C₉H₈O₂), 161 (C₁₀H₉O₂), and 194 (C₁₀H₁₀O₄).

Such comparisons are possible, since it is known that variation in substituents on ring A of pterocarpans does not affect the n.m.r. of ring D protons.7

¹ K. O. Müller and H. Börger, Arb. Biol. Anst. (Reichsaust) Berlin, 1941, 23, 189.

² (a) D. G. Smith, A. G. McInnes, V. J. Higgins, and R. L. ²⁴ (a) D. G. Smith, A. G. McLinnes, V. J. Higgins, and R. L. Millar, *Physiol. Plant Pathol.*, 1971, **1**, 41; (b) D. R. Perrin and W. Bottomley, *J. Amer. Chem. Soc.*, 1962, **84**, 1919; (c) A. Stoessl, *Canad. J. Biochem.*, **50**, 107; (d) D. R. Perrin, *Tetrahedron Letters*, 1964, 29; (e) D. R. Perrin, C. P. Whittle, and T. J. Batterham, *ibid.*, 1972, 1673; (f) J. J. Sims, N. T. Keen, and V. K. Honwad, *Phytochemistry*, 1972, **11**, 827. demonstrated that the total substituent mass can be correctly divided between the rings on the basis of mass spectral evidence. For example, if a pterocarpan is known to possess hydroxy-dimethoxy-substitution, distinction can be made between structures (i) requiring the methoxy-groups on one ring and the hydroxy-group



on the other and (ii) requiring hydroxy-methoxysubstitution on one ring and a methoxy-group on the other.

In the mass spectrum of compound (1) three of the four possible fragment ions indicating methoxy- and hydroxy-dimethoxy-ring substitution are present.[†] Fragments consistent with any other substitution pattern are lacking. In the n.m.r. spectrum signals for three of the four aromatic protons appear as an ABX system. These were assigned to C-7, C-8, and C-10 on the basis of correspondence of the observed chemical shifts and coupling constants with those of 9-methoxypterocarpans.^{2a, 3a, 5, \ddagger} The alternative assignment of the ABX protons to C-1, C-2, and C-4 was ruled out by hydrogenolysis of compound (1) to form the corresponding isoflavandiol (5). The major mass spectral fragments of the isoflavandiol (5) (Scheme) confirm that ³ (a) A. Pelter and P. I. Amenechi, J. Chem. Soc. (C), 1969, 887; (b) A. Pelter, P. Stainton, and M. Barber, J. Heterocyclic

Chem., 1965, 2, 262.

⁴ M. Nakayama, S. Eguchi, A. Matsuo, S. Hayashi, S. Hishida, ⁵ K. G. R. Pachler and W. G. E. Underwood, *Tetrahedron*,

1967, 23, 1817.

⁶ L. H. Briggs, L. D. Colebrook, H. M. Fales, and W. C. Wildman, Analyt. Chem., 1957, 29, 904.

⁷ S. H. Harper, A. D. Kemp, W. G. E. Underwood, and R. V. M. Campbell, *J. Chem. Soc.* (C), 1969, 1109.

ring D of the parent pterocarpan is monomethoxysubstituted.



The problem was thus reduced to determining the positions of the ring-A substituents. Compound (1) reacted positively with 2,6-dichlorobenzoquinone 4-chloroimine (Gibbs reagent),⁸ and thus the lone ring-A proton is *para* to the hydroxy-group. This places the methoxygroups at C-2 and C-3. The position of the proton was determined by comparison of its n.m.r. signal with the calculated chemical shift values of Ballantine and Pillinger,⁹ as amended by Pelter and Amenechi.^{3a} The



possible positions of the proton are shown in formulae (7) and (8); the numerals in parentheses are the calculated values. The observed signal at δ 6.61 places the proton at C-1 and indicates that compound (1) is 4hydroxy-2,3,9-trimethoxypterocarpan.

Compound (2), $C_{17}H_{16}O_5$ (M⁺ 300), contains one less methoxy-group than compound (1). It formed a single methylation product $(M^+ 314)$, a single acetylation product $(M^+ 342)$, and possesses an i.r. spectrum similar to that of compound (1). Analysis of the mass spectrum indicated hydroxymethoxy- and methoxy-substitution; signals corresponding to these substituents appear in the n.m.r. spectrum. A negative Gibbs test indicated substitution *para* to the aromatic hydroxy-group.

The aromatic region of the n.m.r. spectrum contains signals for five protons. Three of these appear as an ABX system which is nearly identical to that of compound (1). Assignment of these protons to C-7, C-8, and C-10 (rather than C-1, C-2, and C-4) was verified ⁸ F. E. King, T. J. King, and L. C. Manning, J. Chem. Soc.,

J. A. Ballantine and C. T. Pillinger, Tetrahedron, 1967, 23, 1691.

¹⁰ G. J. H. Rall, J. P. Englebrecht, and A. J. Brink, Tetrahedron, 1970, **26**, 5007.

by preparation of the isoflavandiol (6). Its mass spectrum possesses ring-B fragments analogous to those of the isoflavandiol (5) (Scheme).

The two remaining aromatic singlets could only have arisen from para-protons. Comparison with the published spectra of 2,3-oxygenated pterocarpans¹⁰ allowed us to assign the signals to H-4 and H-1, respectively, but the data did not permit us to distinguish between 2-hydroxy-3-methoxy- and 3-hydroxy-2-methoxy-substitution. Choice was made by comparison of the H-1 and H-4 signals of compound (2) with those of its acetate. Data from other pterocarpans indicate that acetylation of an aromatic hydroxy-group induces a greater downfield shift of ortho- than of meta-protons. 3a, 10, 11 The phenomenon also occurs with other phenols and their acetates 12 and has been utilized in the structure determination of pterocarpans.¹⁰ The 2-hydroxy-structure requires a greater shift for the H-l signal of the acetate and the 3-hydroxy-structure requires a greater shift for the H-4 signal of the acetate. The greater observed downfield shift for H-4 (0.16 p.p.m.; cf. 0.12 p.p.m. for H-1) is evidence that compound (2) is 3-hydroxy-2,9dimethoxypterocarpan.

Compound (3), $C_{18}H_{18}O_5$ (M⁺ 314), did not form methylation or acetylation products. Its u.v., n.m.r., and mass spectra are identical with those of the mono-O-methyl derivative of compound (2), and thus compound (3) is 2,3,9-trimethoxypterocarpan.¹³

Large negative optical rotations ($[\alpha]_{D}^{23}$ -185, -297, and -228°) were recorded for compounds (1)-(3) respectively, indicating that the compounds possess the (6aR, 11aR) absolute configuration, which is common to laevorotatory pterocarpans.¹⁴

Preliminary studies indicate that the pterocarpans described here exhibit antifungal activities comparable to those of other known pterocarpans.¹⁵ Results of these studies will be published elsewhere.

EXPERIMENTAL

M.p.s were determined with a Fisher-Johns apparatus. U.v. spectra were recorded with a Bausch and Lomb Spectronic UV200 spectrophotometer, i.r. spectra with a Perkin-Elmer 421 instrument (for films on NaCl crystals), and mass spectra with an A.E.I. MS9 instrument (A.E.I. MS902 for high resolution measurements). N.m.r. spectra of dilute solutions in CDCl₃ were recorded at 90 MHz with a Bruker HX90 instrument utilizing the fast Fourier transform technique. Optical rotation measurements were obtained with a Perkin-Elmer 141 polarimeter. T.l.c. was performed with Polygram Sil G-UV₂₅₄ (Brinkmann Instruments) or silica gel with fluorescent indicator 13181 (Eastman Kodak Co.).

11 (a) K. Fukui, M. Nakayama, and T. Harano, Bull. Chem. Soc. Japan, 1960, **42**, 233; (b) L. Farkas, A. Gottesegen, and M. Nógrádi, J.C.S. Perkin I, 1974, 305; (c) K. K. Purushothaman, V. N. Kishore, V. Narayanaswami, and J. D. Connolly, J. Chem. Soc. (C), 1971, 2420.

¹² R. G. Cooke and I. D. Rae, Austral. J. Chem., 1964, 17, 379. ¹³ V. K. Kalra, A. S. Kukla, and T. R. Seshadri, Indian J. Chem., 1967, 5, 607. ¹⁴ S. Ito, Y. Fujise, and A. Mori, Chem. Comm., 1965, 595.

¹⁵ D. R. Perrin and I. A. M. Cruickshank, Phytochemistry, 1969, **8**, 971.

Production and Extraction of Diseased Epicotyls.-Seeds of Pisum sativum were planted in flats of steamed soil, which were kept in a glasshouse. After 7 days a spore suspension of Fusarium solani f. sp. pisi was poured onto the flats of seedlings. The dark lesions which soon appeared on the epicotyls were allowed to enlarge for 15 to 20 days. The plants were then sacrificed, and the excised epicotyls were comminuted in 95% ethanol (4 v/w). After filtration one volume of water was added, and the ethanol was removed under reduced pressure. The aqueous fraction was extracted with chloroform $(2 \times 4 \text{ volumes})$, and the chloroform was evaporated off under reduced pressure. The residue was redissolved in a small amount of chloroform and subjected to t.l.c. [PhH-EtOAc-PrⁱOH (90:10:1)]. U.v.-absorbing bands 1-4, at $R_F 0.31$, 0.39, 0.52, and 0.63, respectively, were resolved; several other bands at lower $R_{\rm F}$ values were not investigated. Band 2 was identified as 6a-hydroxy-3-methoxy-8,9-methylenedioxypterocarpan (pisatin)^{2b} and was not further examined. Further t.l.c. [PhMe-EtOAc (8:1)] of bands 1, 3, and 4 yielded, respectively, compounds (1) $(R_{\rm F} \ 0.48)$, (2) (0.68), and (3) (0.84). Yields varied considerably from batch to batch of epicotyls. Typically, 410 g of epicotyls produced 15.4 mg of compound (1), 7.7 mg of (2), 2.6 mg of (3), and 25.0 mg of pisatin.

4-Hydroxy-2,3,9-trimethoxypterocarpan (6a,11a-Dihydro-2,3,9-trimethoxy-6H-benzofuro[3,2-c][1]benzopyran-4-ol) (1). —The compound crystallized as needles from aqueous acetone; m.p. 141—145°; $[\alpha]_{D}^{23}$ —185° (EtOH); ν_{max} 3420 (OH), 1495, 1595, 1620 (aryl), 1045, 1140, 1275, 1335, and 1470 cm⁻¹ (aryl ether); λ_{max} (EtOH) 287 and 290sh nm (log ε 3·82 and 3·80) (Found: M^+ , 330·1120. C₁₈H₁₈O₆ requires M, 330·1103); m/e 330 (100%), 329 (11), 316 (11), 315 (35), 283 (13), 182 (18), 179 (13), 157·5 (13), 148 (15), and 137 (10); δ 7·13 (1H, d, J 9 Hz, H-7), 6·61 (1H, s, H-1), 6·44 (1H, d, J 2·5 Hz, H-10), 6·44 (1H, q, J 9 and 2·5 Hz, H-8), 5·63br (1H, s, OH), and 3·88, 3·84, and 3·73 (9H, 3 × s, OMe).

4',6,7-Trimethoxyisoflavan-2',8-diol (5).—The pterocarpan (1) (0.8 mg) was dissolved in ethanol (9 ml) and 0.2Nhydrochloric acid (1 ml). 10% Palladium-charcoal (5 mg) was added. Hydrogenation at room temperature and pressure was stopped after 30 min. The residue left after removal of solvent and catalyst was subjected to t.l.c. $[\text{Et}_2\text{O-C}_6\text{H}_{14}$ (5:1)]. The major *product* (R_F 0.22) showed λ_{max} (EtOH) 282sh and 286 nm, and reacted positively with FeCl₃ and 2,6-dichlorobenzoquinone 4-chloroimine (Found: M^+ , 332·1253. C₁₈H₂₀O₆ requires M, 332·1260); m/e 332 (100%), 196 (16), 184 (16), 183 (32), 182 (28), 180 (16), 150 (48), 149 (68), 137 (72), 121 (24).

3-Hydroxy-2,9-dimethoxypterocarpan (2).—The compound crystallized as plates from acetone; m.p. 146—148°; $[\alpha]_{\rm D}^{23}$ -297° (EtOH); $\nu_{\rm max.}$ 3420 (OH), 1495, 1595, 1620 (aryl), 1030, 1145, 1275, 1340, and 1465 cm⁻¹ (aryl ether); $\lambda_{\rm max.}$ (EtOH) 288sh and 292 nm (log ε 3·98 and 4·01) (Found: M^+ , 300·0988. C₁₇H₁₆O₅ requires M, 300·0993); m/e 300 (100%), 299 (18), 285 (23), 161 (10), 150 (10), 149 (10), and 148 (26); δ 7·12 (1H, d, J 8·8 Hz, H-7), 6·96 (1H, s, H-1), 6·52 (1H, s, H-4), 6·44 (1H, d, J 2·3 Hz, H-10), 6·43 (1H, q, J 8·8 and 2·3 Hz, H-8), 5·89br (1H, s, OH), and 3·90 and 3·77 (6H, 2 × s, OMe).

4',7-Dimethoxyisoflavan-2',6-diol (6).—The pterocarpan (2) (0·29 mg) was hydrogenated as described above. The major product ($R_{\rm F}$ 0·38) showed $\lambda_{\rm max}$. (EtOH) 287 nm, and reacted positively with FeCl₃ and 2,6-dichlorobenzoquinone 4-chloroimine (Found: M^+ , 302·1142. C₁₇H₁₈O₅ requires M, 302·1154); m/e 302 (96%), 166 (18), 165 (29), 164 (32), 154 (14), 153 (46), 151 (21), 150 (100), 149 (25), 138 (14), 137 (54), 133 (29), and 131 (14).

2,3,9-Trimethoxypterocarpan (3).—The compound crystallized as needles from benzene-heptane; m.p. 122—124°; $[a]_{\rm p}^{23}$ —228° (EtOH); $v_{\rm max}$ 1500, 1600, 1620 (aryl), 1030, 1140, 1270, 1345, and 1450 cm⁻¹ (aryl ether); $\lambda_{\rm max}$ (EtOH) 288sh and 292 nm (log ε 3·96 and 3·98) [lit.,¹³ m.p. 121— 122°; $v_{\rm max}$ (KBr) 2985, 1623, 1510, 1418, 1387, 1348, 1269, 1195, 1176, 1081, 1042, 960, and 855 cm⁻¹; $\lambda_{\rm max}$ (MeOH) 290 nm (log ε 3·35)] (Found: M^+ , 314·1133. C₁₈H₁₈O₅ requires M, 314·1154); m/e 314 (100%), 313 (24), 312 (10), 311 (10), 299 (28), 161 (10), and 148 (31); δ 7·12 (1H, d, J 8·8 Hz, H-7), δ ·97 (1H, s, H-1), δ ·47 (1H, s, H-4), δ ·45 (1H, d, J 2·5 Hz, H-10), δ ·44 (1H, q, J 8·8 and 2·5 Hz, H-8), and 3·91, 3·86, and 3·78 (9H, 3 × s, OMe).

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